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## **ViPrimePLUS Trypanosoma equiperdum qPCR Kit**

Quantitative assay for real-time PCR detection of Trypanosoma equiperdum genome

**Product Code: QV3007**  
**Pack Size: 150 reactions**

## INTENDED USE

The ViPrimePLUS *Trypanosoma equiperdum* qPCR Kit is a Taqman probe-based real-time PCR assay for the detection of *Trypanosoma equiperdum* genome in urine, urogenital and vaginal swab. This assay is intended for research use only.

## INTRODUCTION

*Trypanosoma equiperdum* is a protozoan responsible for dourine disease among horses (most susceptible), mules and donkeys. Dourine is the only trypanosomiasis that is not spread via an invertebrate vector, but rather it is transmitted sexually, during coitus. The disease manifests various symptoms. Classically, it is characterized into three stages. The first stage will observed swelling of genitalia, abnormal vaginal discharges, patch discoloration in the vulva and penis. Fever and loss appetite may occur. The second stage follows after 4weeks, where intermittent onset of round urticarial formation appear around the neck, chest and flanks. In the third stage, paralysis of the limbs takes place, thus leading to death though the disease may persist for a year or two. However, most of the infected animals may only developed mild symptoms, especially those from endemic regions. Diagnosis is difficult as the trypanosomes are not present in blood, but in the genital tissue or fluids. Serological test could not differentiate between *T. equiperdum* and other trypanosomes, such as *T. evansi* and *T. brucei*. Thus, molecular diagnosis is a good alternative in terms of sensitivity and specificity.

## PRINCIPLE OF TEST

The kit contains primers and Taqman® probe that target the germline variant surface antigen (VSG-78 BC-3) pseudogene.

Based on the Taqman® probe detection principle, the 5'-reporter dye and 3'-quencher dual-labelled oligonucleotide (Taqman® probe) hybridizes on a specific region within the amplified fragment. During amplification, the probe is cleaved and the reporter dye (fluorophore) is released. The fluorescent signal intensity detected is proportional to the number of amplicons. The Ct value (the cycle at which the rise of fluorescent signal from the baseline is first significant) is used for quantification purposes. Target pathogen amplification is detected using FAM channel.

The kit provides the Internal Extraction Control (IEC) as inhibition control. During nucleic acid extraction protocol, IEC DNA template is added in the lysis stage. An IEC specific primers and probe labelled with a different dye is provided to be run in the same reaction with the pathogen-specific primers and probe mix. The IEC amplicons are detected via VIC/HEX channel at Ct value 28±3 depending on the sample dilution.

A positive control with known copy number is provided for standard curve construction and absolute quantification. It can also be used at a single dilution for qualitative analysis control of the experimental set-up. Extra care must be taken to avoid cross-contamination.

## QUALITY CONTROL

Each lot of ViPrimePLUS *Trypanosoma equiperdum* qPCR Kit has been tested against predetermined specifications to ensure consistent product quality under ISO 9001:2008 – certified Quality Management System.

## SENSITIVITY & SPECIFICITY

The detection limit is tested to 100 copies per reaction. The primers and probe are 100% specific.

## STORAGE & STABILITY

Store at -20°C and avoid light exposure. Stable at -20°C up to the expiry date stated. Keep in aliquot to reduce freeze-thaw cycles.

## LIMITATION OF TEST

For research use only. Not recommended for diagnosis of disease in humans or animals.

Result is dependent on the yield and quality of the nucleic acids extracted from the method of extraction. Thus, it is important to do spectrophotometric and gel analysis on the extracted samples.

## KIT COMPONENTS

|   |                    |
|---|--------------------|
| T. equiperdum Primers and Probe Mix (T. equiperdum PPM)     | Amber Capped Tube  |
| Positive Control  | Pink Capped Tube   |
| Internal Extraction Control Primers and Probe Mix (IEC PPM) | Amber Capped Tube  |
| Internal Extraction Control (IEC DNA)                       | Blue Capped Tube   |
| Nuclease Free Water   | White Capped Tube  |
| Template Preparation Buffer                                 | Yellow Capped Tube |

## SAMPLE MATERIAL

The kit is suitable for DNA extracted by most commercial kits, provided the purity, concentration and integrity are within acceptable range. IEC is provided to eliminate doubts of PCR inhibition. Suitable sample type is blood.

## PROTOCOLS

### Reconstitution of reagents

\*Pulse-spin each tube prior to opening.

| Components                | Volume | Reagents                    |
|---------------------------|--------|-----------------------------|
| T. equiperdum PPM (Amber) | 165µl  | Nuclease Free Water         |
| IEC PPM (Amber)           | 165µl  | Nuclease Free Water         |
| IEC DNA                   | 600µl  | Nuclease Free Water         |
| Positive Control Template | 500µl  | Template Preparation Buffer |



### DNA extraction

Add 4µl of IEC DNA into each sample suspended in the lysis/extraction buffer.

**\*DO NOT** add IEC directly into unprocessed biological sample.



### Real-time PCR reaction set-up

Recommended real-time PCR reaction set-up:

| Reagents  | 1 reaction (µl) |
|---|-----------------|
| 2x PCR Mastermix  | 10              |
| T. equiperdum PPM   | 1               |
| IEC PPM (not required when preparing standards' reaction mix) | 1               |
| Nuclease-free water   | 3               |
| Sample DNA  | 5               |

\*Suggested sample concentration 5-20ng/µl.

\*Adjust nuclease-free water to make up the final reaction volume of 20µl.

\*Prepare an extra reaction to accommodate for pipetting error.

#### Negative control

Set aside one tube/well as negative control where 5µl of nuclease-free water is used as the template. This is also known as no template control.

#### Positive control

For qualitative analysis, set aside one tube as positive control where 5µl of the positive control is used as the template.

#### Positive control (optional)

For quantitative analysis only, prepare 6 serially diluted positive control templates for standard curve construction.

- Pipette 90µl of nuclease-free water into 5 clean microtubes and label 2-6 accordingly.
- Pipette 10µl of Positive Control into tube 2.
- Vortex thoroughly and spin down.
- Change pipette tip and pipette 10µl from tube 2 to tube 3. Vortex and spin down.
- Repeat steps d) to complete the dilution series.
- Pipette 5µl of standard template into each tube/well accordingly.

| Standards               | Copy number/µl  |
|-------------------------|-----------------|
| Tube 1 Positive Control | $2 \times 10^5$ |
| Tube 2                  | $2 \times 10^4$ |
| Tube 3                  | $2 \times 10^3$ |
| Tube 4                  | $2 \times 10^2$ |
| Tube 5                  | 20              |
| Tube 6                  | 2               |

Set the thermal cycler parameters as follows:

| Step              | Time   | Temp | Cycles | Scan |
|-------------------|--------|------|--------|------|
| Enzyme activation | 2mins  | 95°C |        |      |
| Denaturation      | 10secs | 95°C | 50     |      |
| Anneal/Elongation | 1min   | 60°C |        | √*   |

\*FAM or VIC/HEX

## INTERPRETATION OF RESULTS

Pathogen specific amplification signal is detected via FAM channel, while IEC amplification is detected via VIC/HEX channel. The signal is positive if the amplification curve crosses the threshold line. The result is relevant provided both positive and negative controls give valid results.

### Summary of interpretation:

| Target | IEC | Negative Control | Positive Control | Interpretation  |
|--------|-----|------------------|------------------|-----------------|
| +      | +   | -                | +                | Valid, positive |
| +      | -   | -                | +                | Valid, positive |
| -      | +   | -                | +                | Valid, negative |
| -      | -   | -                | -                | Invalid         |
| +      | +   | +                | +                | Invalid         |

### Internal Extraction Control

When used accordingly and assuming 100% extraction efficiency, a Ct value of 28±3 is within normal range. A high Trypanosoma equiperdum genome copy amplification may out compete the IEC amplification. Thus, the latter may not produce an amplification signal. The positive result is still valid in this case.

## TROUBLESHOOTING

| Problem   | Possibility  | Suggestion  |
|---|--|---|
| Negative control / No template control gives positive result                      | Carry over contamination   | Change nuclease-free water. Use fresh aliquots of reagents. Use filtered tips. Load positive control last.    |
| No signal detected from positive control  | Incorrect programming of instrument  | Check program.  |
|   | Reagents expired   | Check the expiry date of reagents before repeat.  |
|   | Storage condition not complying with instructions  | Check storage condition properly and store at correct storage condition to avoid the degradation of reagents. |
| Internal extraction control does not give a signal in apparently negative samples | Pipetting error  | Pipette the correct volume of reagents to reconstitute the components of kit and mix well.                    |
|   | Inhibitors in the samples extracted  | Repeat the extraction.  |
|   | Low recovery of RNA extracted  | Repeat the extraction by enlarge the sample size.   |
| Internal extraction control does not give a signal in apparently negative samples | IEC added directly into unprocessed biological sample – lead to degradation and loss of signal | Add IEC into each sample suspended in the lysis/extraction buffer.  |

## DEVIATION OF MASTERMIX FORMULATION

Manufacturers use varying methods to calibrate a real-time PCR reaction. For this reason, we provide several Mastermix formulations for those platforms.

| Master Mix                   | Compatible Hardware  |
|------------------------------|--|
| Original                     | Biometra qTower, Cepheid SmartCycler®, Eppendorf Mastercycler, Fluidigm BioMark™, Illumina Eco, MJ Chromo4, Opticon, PCRMax Eco™, Roche lightcycler® 480, lightcycler® LC96 and lightcycler® Nano Platforms, RotorGene, Thermo PikoReal™ |
| Low Rox (-LR)                | Applied Biosystems 7500 and 7500 FAST platform, QuantStudio™, ViiA7.   |
| Rox (-R)                     | Applied Biosystems 7000, 7300, 7700, 7900 and 7900HT FAST platforms, GeneAmp® 5700, StepOne™, StepOne™ PLUS  |
| iCycler platform (-iC)       | BioRad iCycler, IQ4 AND IQ5 platforms  |
| Stratagene platform (-SG)    | Stratagene MX, MX4000P®, MX3000P® and MX3005® platforms  |
| Capillary lightcyclers (-CL) | Roche Capillary Lightcycler 1.0-2.0.   |

## WARRANTY & LIMITED LIABILITY

The performance characteristics stated were obtained using the assay procedure in this insert. Failure to comply with the instructions may derive inaccurate results. In such event, manufacturer disclaims all warranty expressed, implied or statutory including the implied warranty of merchantability and the fitness of use.

The manufacturer will not be liable for any damage caused by misuse, improper handling and storage; non-compliance with precautions and procedures, and damages caused by events occurring after the product is released.